

TITLE:

PROCEDURE FOR DETERMINATION OF BROMINE AND CHLORINE IN SOLUTIONS APPLIED TO MEAT AND POULTRY PRODUCTS (Modified DPD Method)

PURPOSE:

This document is to be used by any lab personnel to determine the amount of bromine and chlorine in activated HOBr solutions using the HACH DR/890 Colorimeter or equivalent. This test is performed on the solution applied to the meat and poultry products e.g. the carcass wash, head wash or OLR cabinet. It is recommended that this procedure be followed at least on a weekly basis (or at each plant's discretion, more frequently) because this method can distinguish between hypobromous acid and chlorine bleach.

EQUIPMENT / REAGENTS:

- HACH DR/890 Colorimeter- Catalog number 48470-00 (or equivalent)
- or: HACH Colorimeter 2 Catalog number 58700-00 (or equivalent)
- Distilled water
- 10 % Glycine solution (Enviro Tech #D-10015-03) or equivalent.
- DPD FREE Chlorine Reagent Pillow Packets (for 10 mL)- HACH # 21055-69
- Potassium Iodide Crystals (KI) – SHAPE Products catalog #8118 (or equivalent)
- 100 ml graduated cylinder
- 5 ml syringe
- 1 ml syringe

PROCEDURE:

Before testing make sure the instrument is in the low (LO) range mode by checking that the display reads to the hundredths (0.00).

The following analytical procedure should be performed with 2 minutes of drawing the sample at the carcass wash.

1. Into the 100 ml graduated cylinder, add 97 ml of distilled water. Add 2 ml of 10 % glycine solution using the 5 ml syringe, cover and invert the cylinder to mix, then add 1 ml of test solution using the 1 ml syringe. (The glycine masks the chlorine concentration at this point). Invert the cylinder several times to mix. The markings on the 1 ml syringe will rub off very easily, so it is recommended to score the 1 ml marking with a knife or scissors.



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Facility: Modesto

Approval Name & Signature:
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Approval Title: VP Technology

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Revised Section(s):
IV. Time limitation between drawing sample and performing analysis

Without a yellow control stamp to the right of this statement, this procedure is a draft. A draft or an uncontrolled copy cannot be used to manage a process or task.

2. Fill both 10 ml sample cells with 10 mls of the sample prepared in Step 1. Designate one of these to be the blank and the other to be the prepared sample. Make sure the cells are not wet and they are free of fingerprints or smudges.
3. Cap the blank cell and place it in the cell holder with the diamond mark facing you. Cover the cell compartment and press ZERO. The instrument will display 0.00. Remove the blank.
4. Add the contents of one DPD **FREE** Chlorine pillow packet to the prepared sample cell. Cap and shake vigorously. A pink color will develop.
5. Place the sample cell in the compartment with the diamond mark facing you, close the cover and press READ.
6. The instrument display will show "--" followed by the results in ppm total chlorine. Call this reading "B" for Bromine.
7. Note: if the instrument display is blinking, the sample concentration is too high and needs to be diluted further before resuming with step #8.
8. Remove the sample cell from the compartment and add a small amount of potassium iodide (KI) crystals (two crystals) to the sample cell still containing the sample. Vigorously shake for 15 seconds, this step allows the glycine-combined chlorine to react with the potassium iodide (KI).
9. Place the sample cell in the compartment with the diamond mark facing you, close the cover and press READ. This reading is ppm Total Halogen. Call this reading TH.

Ideally, results from step 6 and step 9 should be the same, $\pm 5\%$: TH = B.

If not, then some of the halogen is present as Chlorine (Cl)*:
Therefore,

$$\text{ppm Cl} = \text{TH} - \text{B}$$

Calculations:

$$\text{ppm as Bromine} = \text{B} \times 2.25 \times 100 \text{ (dilution factor)}$$

$$\text{ppm as Chlorine} = \text{TH} - \text{B} \times 100 \text{ (dilution factor)}$$

**Reminders:

- (1) Multiply colorimeter reading by dilution factor used.
- (2) Sample cell and cell cap must be washed thoroughly between replicates to remove KI residues, which will interfere with the accuracy of the subsequent samples.
- (3) Distilled water is recommended as it has been found to be less susceptible to interferences that Reverse Osmosis (RO) or De-ionized (DI) water